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# Exploring odontocete depredation rates in a pelagic longline fishery

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## Abstract

Several odontocete species depredate catch and bait from fishing gear, resulting in their bycatch and causing substantial economic costs. There are no known mitigation methods for odontocete depredation in pelagic longline fisheries that are effective, do not harm odontocetes and are commercially viable. Understanding odontocetes' depredation strategies can contribute to mitigating this human-wildlife conflict. Using observer data from the Hawaiibased tuna longline fishery, this study summarized teleost and elasmobranch species-specific mean posterior odontocete depredation rates using a simple Bayesian binomial likelihood estimator with a Bayes-Laplace prior. Depredation rates of species with sufficient sample sizes ranged from a high of 1.2% (1.1 to 1.3 95% highest posterior density interval or HDI) for shortbill spearfish to a low of 0.002% (0.001 to 0.003 95% HDI) for blue shark. Depredation of catch is a rare event in this fishery, occurring in about 6% of sets. When depredation did occur, most frequently odontocetes depredated a small proportion of the catch, however, there was large variability in depredation rates between teleost species. For example, bigeve tuna was two times more likely to be depredated than vellowfin tuna (odds ratio = 2.03, 95% CI: 1.8-2.3, P<0.0001). For sets with depredation, 10% and 2% of sets had depredation of over half of the captured bigeye tuna and combined teleosts, respectively. All elasmobranch species had relatively low depredation rates, where only 7 of almost 0.5M captured elasmobranchs were depredated. Odontocetes selectively depredate a subset of the teleost species captured within sets, possibly based on net energy value, chemical, visual, acoustic and textural characteristics and body size, but not median length, which was found to be unrelated to depredation rate (Pearson's r = 0.14, 95% CI: -0.26 to 0.50, p = 0.49). Study findings provide evidence to support the identification and innovation of effective and commercially viable methods to mitigate odontocete depredation and bycatch.

## 1. Introduction

Marine mammals occupy a range of trophic levels and some species have important roles in regulating the structure, function, dynamics and state of marine communities and ecosystems [1–6]. Large changes in their abundance can therefore cause profound community- and

authors are prevented from making the U.S. government observer program data publicly available. All other datasets used in this study are open access and citations to these datasets are included in the article.

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ecosystem-level consequences that are protracted or irreversible. Some marine mammals, including sirenians and most toothed whales, are extremely sensitive to anthropogenic mortality due to having relatively low rates of potential population growth and other intrinsic life history characteristics, small population sizes, and extrinsic environmental variables such as narrow habitat requirements for foraging and breeding [7–9].

Fisheries bycatch in both in-use and derelict (abandoned, lost and discarded) fishing gear is the largest threat to many populations of marine megafauna, including marine mammals, and is an obstacle to sustainable seafood production [9–12]. Of the range of anthropogenic hazards faced by marine mammals, bycatch affects the greatest number of species, contributed to the presumed extinction of the baiji (Yangtze River dolphin, *Lipotes vexillifer*), and has contributed to reducing the abundance of several populations and species of cetaceans and pinnipeds to critical levels [9, 13, 14]. Furthermore, marine mammal depredation (removal and damage of catch and bait from fishing gear) causes substantial direct and indirect socioeconomic costs [15, 16]. Depredation can also result in fishers deliberately injuring and killing marine mammals, and alters marine mammals' foraging behavior, distribution, diet and demographics [7, 15–19].

Several species of odontocetes (Odontoceti, toothed whales), including the false killer whale (Pseudorca crassidens), short-finned pilot whale (Globicephala macrorhynchus) and killer whale (Orcinus orca), depredate catch and bait in pelagic longline fisheries, which can result in their bycatch by becoming hooked or entangled in line [20, 21]. A U.S. central north Pacific Ocean tuna longline fishery with vessels based primarily from Hawaii experiences odontocete depredation and bycatch primarily by false killer whales [22]. Under the U.S. Marine Mammal Protection Act, a Take Reduction Team was established in 2010 because the five-year running average of the estimated annual false killer whale mortalities and series injuries (M&SI) exceeded a threshold Potential Biological Removal (PBR) level. The team was tasked with reducing M&SI to below PBR within six months and to a level approaching zero, defined as <10% of PBR, within five years, and was further tasked with having no increase in M&SIs for a high seas false killer whale stock [23]. To date, existing measures recommended by the team and adopted by the US fisheries management authority ('weak' circle hooks, permanent static closed area, fleetwide bycatch cap that triggers an area closure, prescribed handling and release practices [23]) have not achieved <10% of PBR, and M&SIs on the high seas have increased [22]. More effective false killer whale depredation and bycatch mitigation methods are needed.

Unlike for some other threatened taxonomic groups exposed to bycatch in pelagic longline fisheries, there are no known odontocete methods across the tiers of a sequential bycatch mitigation hierarchy (avoid, minimize, remediate and offset) that are: (1) effective, (2) do not harm odontocetes, and (3) have acceptable costs to commercial viability (economic viability, practicality and crew safety) [15, 21, 24, 25]. There is low strength of evidence of the efficacy and commercial viability of bycatch and depredation mitigation approaches for odontocetes in pelagic longline gear. This includes mitigation approaches such as spatiotemporal fisheries management, including quasi-real time dynamic (fleet communication, move-on rules, habitat suitability modelling, real-time acoustic tracking) and static approaches [9, 15, 22, 24, 26, 27], input controls such as limits on soak and haul duration [24, 28], and output controls such as bycatch thresholds [29]. The same applies to gear technology approaches such as:

- Catch encasement to physically protect catch from depredation, and visual and acoustic camouflage of target catch such as by using bubble screens or knots [24, 30, 31];
- Mainline length limits, hookless sections of mainline, hookless sets, and set geometry [32];

- Active acoustic deterrents and decoys, including pingers, acoustic harassment devices, decoy vessels, broadcasts of decoy fishing vessel acoustic cues, broadcasts of killer whale and other predator sounds, masked or disrupted odontocete returning echolocation [21, 24, 33–36];
- Passive acoustic decoys and deterrents such as the incorporation of objects into gear that: simulate the acoustic target strength of odontocete-depredated catch, simulate the target strength of species that odontocetes avoid depredating, are perceived as unusual prey, or interfere with echolocation [24, 37];
- Acoustic masking, including quieter vessels; bubble screen to reduce the dissemination of vessel sounds; broadcast of sounds to conceal the sounds of the vessel gear, setting and hauling; not remaining near the gear after it is set; and minimizing shifting in and out of gear [20];
- Artificial bait [38];
- Weak hooks. While there is no evidence of weak hooks causing reduced odontocete bycatch rates, mechanistic studies found that wire diameter was an informative predictor of the force required to straighten hooks [39–41] and during at-sea experiments weak hooks were observed to straightened more often than control hooks with a wider hook wire diameter [39, 40]; and
- Chemical deterrents [20, 24, 42].

There is also weak evidence of the effect of prescribed pelagic longline handling and release practices on cetacean post-release mortality rates. For instance, estimates of survival probability made by the US Government are based largely on expert opinion [22, 43]. Therefore, evidence is needed to support the identification and development of effective and commercially viable approaches to mitigate odontocete depredation and bycatch rates in pelagic longline fisheries.

Understanding the depredation strategies of odontocetes that are susceptible to fisheries bycatch can contribute to mitigating this human-wildlife conflict. Knowledge of species-specific odontocete depredation rates might contribute to the development of promising, new mitigation methods, such as geospatial, depth and temporal separation of predictable catch rate hotspots of species with both low commercial value and high odontocete depredation rates and hotspots of catch rates of principal market species, and passive acoustic decoys, deterrents and echolocation disruptors. This study analyzed observer data from the Hawaii tuna longline fishery to estimate teleost and elasmobranch species-specific mean posterior marine mammal depredation rates using a Bayesian binomial likelihood estimator with a Bayes-Laplace prior, expanding upon previous assessments [44, 45]. The study also assessed the correlation between species-specific depredation rates and length. Study findings provide evidence to support the innovation of effective and commercially viable methods to reduce the risk of odontocete depredation and bycatch in pelagic longline fisheries.

#### 2. Methods

Data on species-specific catch, presence/absence of marine mammal depredation and length were obtained from the U.S. government observer program database for the Hawaii pelagic longline fishery. Observer data fields and data collection protocols are defined in NMFS [46]. The study period was from 15 August 2003, the date when observers began to record damage to the catch, through 26 August 2023, the most recent available record. The study sample included 179,853,812 hooks deployed in 73,217 sets within 5,445 trips by 194 vessels. Records were included in the study sample to estimate species-specific odontocete depredation rates

for fish species, both teleosts and elasmobranchs, that were identified to the species level, and had >100 capture records where the observer determined whether the catch had been depredated by a marine mammal, Thus, the study sample excludes catch records where either the observer was unable to determine whether there was damage to the catch or observed damage but could not determine the source. Length measurement methods of eye-to-fork was used for billfishes and fork length for all other species, as these produced the largest sample sizes for species within these groups.

The expected depredation rate (or proportion) for each of the 50 species was estimated using a Bayesian binomial likelihood estimator that here accounts for the zero recorded depredation events [47]. A simple binomial estimator is the most appropriate likelihood for these data that comprise the number of depredated observations and the number of not-depredated observations per set (or "successes" and "failures")—see Lin and Chu [48] for further discussion. Moreover, 40% of the 35 species-specific records had zero depredations per set and hence the use of a Bayes-Laplace prior to account for those zero observations explicitly when using a binomial estimator ([47]; and see Gilman et al. [49] for an example for shark at-vessel mortality rates). The mean posterior rate and a 95% highest posterior density interval (HDI) was summarized by sampling from a binomial likelihood with a Bayes-Laplace prior [47] using the binom package for R [50] as proposed elsewhere by Roda et al. [51] and Gilman et al. [49]—rather than just using the raw study-specific summaries.

The relationship between predicted mean depredation rates and median lengths was assessed for teleosts with > 1,000 capture records by calculating the Pearson product-moment correlation coefficient [52] using the corr() and cor.mtest () functions in the corrplot R package [53]. There were insufficient sample sizes available to estimate expected marine mammal depredation rates by length class within species (of 1,038,586 species-level catch records with length values, only 44 were identified as having marine mammal depredation)–observers are typically unable to make common length measurements (total length, eye-to-fork length, fork length) of catch depredated by marine mammals because the depredated catch tends to be retrieved with only the fish head up to the gills, or just the lips and upper jaw of the fish remaining [15, 16]. In other words, we were able to conduct analyses to determine whether catch with larger median lengths had different expected mean odontocete depredation rates than species with smaller median lengths, but not on whether larger individuals within a species had a different odontocete depredation rate than smaller individuals of that species.

The proportion of total sets with odontocete depredation of  $\geq 1$  fish was determined. The proportion of the total catch of bigeye tunas (*Thunnus obesus*), yellowfin tunas (*T. albacares*) and combined teleosts that had odontocete depredation was determined for (1) sets with observed odontocete depredation of  $\geq 1$  fish, and (2) sets with observed odontocete depredation rate for bigeye tuna, yellowfin tuna or teleost. We then also compared the depredation rate for bigeye and yellowfin tunas, the principal target species of this fishery, using the odds ratio [54] calculated by conditional maximum likelihood or Fishers exact procedure using the epitools R package [55].

#### 3. Results

Table 1 presents the mean posterior depredation rates (percent of the catch that was depredated by marine mammals) with 95% HDIs and median and mean lengths. For species with >1,000 capture records, depredation rates ranged from a high of ca. 1.2% (1.1 to 1.3 95% HDI) for shortbill spearfish to a low of 0.002% (0.001 to 0.003 95% HDI) for blue shark. Bigeye tuna was at least two times more likely to be depredated than yellowfin tuna (odds ratio = 2.03, 95% CI: 1.8–2.3, P<0.0001). Elasmobranchs had extremely low marine mammal depredation rates, Table 1. Study sample sizes, mean posterior depredation rates and 95% highest posterior density intervals (HDIs), and length summaries for fish species with >100catch records for which an observer determined whether there was marine mammal depredation in the Hawaii longline tuna fishery (2003–2023). There were179,853,812 observed hooks in 73,217 sets. Lengths are eye-to-fork for billfishes and fork length for all other species. Sorted from high to low mean depredation rate within 5 taxonomic groups.

Scientific name	Common name		% depredated			No. depredated	Length (cm)		
		mean	lower HDI	upper HDI			N	Median	Mean
Tunas									
Katsuwonus pelamis	Skipjack tuna	0.469	0.433	0.505	136,931	641	51,940	71	70
Thunnus obesus	Bigeye tuna	0.292	0.280	0.305	735,284	2,148	288,006	113	112
Thunnus alalunga	Albacore tuna	0.167	0.134	0.202	55,066	91	22,272	101	100
Thunnus albacares	Yellowfin tuna	0.144	0.127	0.162	177,441	255	68,398	117	110
Billfishes									
Tetrapturus angustirostris	Shortbill spearfish	1.172	1.088	1.256	62,722	734	24,320	134	134
Xiphias gladius	Swordfish	0.466	0.389	0.545	29,386	136	10,643	93	109
Kajikia audax	Striped marlin	0.387	0.338	0.436	61,259	236	26,001	137	134
Istiophorus platypterus	Indo-Pacific sailfish	0.263	0.077	0.475	2,283	5	800	145	141
Makaira nigricans	Blue marlin	0.229	0.165	0.297	20,055	45	7,646	161	166
Other teleosts									
Elagatis bipinnulata	Rainbow runner	1.775	0.185	3.764	167	2	34	73	75
Acanthocybium solandri	Wahoo	1.060	0.992	1.128	87,450	926	24,876	127	126
Uraspis secunda	Cottonmouth jack	0.952	0	2.839	103	0	28	32	31
Brama japonica	Pacific pomfret	0.877	0	2.616	112	0	74	33	35
Lampris megalopsis	Bigeye Pacific opah	0.579	0.524	0.635	71,311	412	27,415	101	100
Assurger anzac	Razorback scabbardfish	0.386	0	1.154	257	0	77	224	218
Coryphaena hippurus	Dolphinfish	0.332	0.310	0.353	278,395	922	77,893	85	86
Masturus lanceolatus	Sharptail mola	0.331	0	0.990	300	0	16	94	93
Trachipterus fukuzakii	Tapertail ribbonfish	0.253	0.005	0.601	790	1	187	194	190
Nesiarchus nasutus	Black gemfish	0.250	0	0.748	398	0	144	70	67
Ranzania laevis	Slender mola	0.218	0.099	0.349	5,037	10	1,196	50	51
Mola mola	Common mola	0.211	0	0.630	473	0	4	98	113
Promethichthys prometheus	Roudi's escolar	0.112	0	0.336	889	0	253	64	65
Sphyraena barracuda	Great barracuda	0.105	0.045	0.172	9,488	9	2,762	99	101
Scombrolabrax heterolepis	Longfin escolar	0.105	0.039	0.179	7,595	7	1,889	24	24
Omosudis lowii	Hammerjaw	0.103	0	0.309	968	0	236	27	28
Eumegistus illustris	Lustrous pomfret	0.095	0	0.284	1,054	0	439	49	51
Taractichthys steindachneri	Sickle pomfret	0.085	0.074	0.097	230,917	196	96,573	60	58
Lepidocybium flavobrunneum	Escolar	0.079	0.067	0.093	179,932	142	47,235	76	76
Lagocephalus lagocephalus	Pelagic puffer	0.066	0	0.198	1,510	0	369	46	45
Gempylus serpens	Snake mackerel	0.062	0.052	0.071	256,820	157	63,820	99	101
Taractes asper	Rough pomfret	0.055	0	0.165	1,815	0	611	50	53
Coryphaena equiselis	Pompano dolphinfish	0.049	0	0.146	2,051	0	666	57	58
Alepisaurus ferox	Longnose lancetfish	0.032	0.028	0.035	855,920	270	179,579	113	105
Ruvettus pretiosus	Oilfish	0.032	0.001	0.076	6,228	1	1,686	72	86
Taractes rubescens	Dagger pomfret	0.012	0	0.028	17,135	1	5,676	67	64
Rays									
Mobula mobular	Devil ray	0.261	0	0.781	381	0	0	NA	NA
Pteroplatytrygon violacea	Pelagic stingray	0.004	0	0.012	24,375	0	27	52	59
Sharks									
Carcharhinus galapagensis	Galapagos shark	0.926	0	2.761	106	0	1	115	115
Carcharhinus plumbeus	Sandbar shark	0.595	0	1.778	166	0	5	109	121

(Continued)

Scientific name	Common name	% depredated			No. caught	No. depredated	Length (cm)		
		mean	lower HDI	upper HDI			Ν	Median	Mean
Sphyrna zygaena	Smooth hammerhead shark	0.301	0	0.901	330	0	13	197	191
Isurus paucus	Longfin mako shark	0.112	0	0.334	894	0	95	156	158
Alopias pelagicus	Pelagic thresher shark	0.099	0	0.297	1,006	0	87	125	122
Zameus squamulosus	Velvet dogfish	0.020	0	0.059	5,106	0	1,013	69	68
Carcharhinus falciformis	Silky shark	0.019	0	0.058	5,153	0	270	108	114
Pseudocarcharias kamoharai	Crocodile shark	0.018	0	0.053	5,618	0	642	85	84
Carcharhinus longimanus	Oceanic whitetip shark	0.015	0	0.046	6,475	0	363	106	107
Alopias superciliosus	Bigeye thresher shark	0.009	0.001	0.018	34,728	2	596	156	157
Isurus oxyrinchus	Shortfin mako shark	0.006	0	0.018	17,072	0	1,012	185	186
Prionace glauca	Blue shark	0.002	0.001	0.003	340,967	5	566	175	176

#### Table 1. (Continued)

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where of a total of 442,377 captured elasmobranchs identified to the species level and for which the observer determined whether marine mammal depredation occurred, 7 were depredated (2 bigeye thresher sharks and 5 blue sharks) (Table 1).

Fig 1 presents the predicted mean depredation rate (proportion of the catch with marine mammal depredation) and 95% HDIs for the four species of tunas included in Table 1, the five species of billfishes included in Table 1, and the six species of other teleosts with the highest predicted mean depredation rates and with sufficient sample sizes of >1,000 capture records. Species are sorted from lowest to highest depredation rate within each of the three taxonomic groups.

Consistent with this species-selective depredation, there was high variability in the speciesspecific set-level marine mammal depredation rates. For example, of sets with  $\geq 1$  depredated bigeye tuna, the main target species of this fishery, 94% had depredation of over half of the captured bigeye tuna. This rate was 42% for yellowfin tuna (i.e., of sets with  $\geq 1$  depredated yellowfin tuna, 42% had depredation of over half of the captured yellowfin tuna), and 2% for combined teleost species. Similarly, for sets with depredation of  $\geq 1$  fish of any species, 10%, 4% and 2% of these sets had depredation of over half of the captured bigeye tuna, yellowfin tuna and combined teleosts, respectively. For sets with depredation of  $\geq 1$  fish of any species, 4%, 2% and 0.1% of these sets had odontocete depredation of all the captured bigeye tuna, yellowfin tuna and combined teleosts, respectively. Of the sets in the study sample, 6.2% had observed odontocete depredation of  $\geq 1$  fish.

There was no significant relationship between median length and mean depredation rate: Pearson's r = 0.14 (95% CI: -0.26 to 0.50), p = 0.49. The species-specific median length of the catch was not an informative predictor of odontocete depredation rate of teleosts.

### 4. Discussion and conclusions

Marine mammal depredation is a rare event in this fishery, occurring in about 6% of sets. When depredation did occur, most frequently a small proportion of the total catch was depredated. However, there was large variability in odontocete depredation rates between teleost species, while all species of captured elasmobranchs had negligible odontocete depredation. This is consistent with previous studies of the observer program database for the Hawaii tuna longline fishery [44, 45]. Findings suggest that odontocetes selectively depredate a subset of the teleost species captured within sets. Odontocetes are not indiscriminately depredating the catch down the line, but are selectively depredating certain teleost species.



expected species-specific marine mammal depredation rate

proportion of catch depredated

Fig 1. Mean posterior depredation rates (proportion of the number captured that was depredated by odontocetes) and 95% highest posterior density intervals (HDIs) for tunas, billfishes and six other teleost species with the highest predicted mean depredation rates and with >1,000 capture records in the Hawaii tuna longline fishery (2003–2023). Dot size is proportional to catch for that species relative to the catch for all three taxonomic groupings.

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Different species of pelagic marine predators, and sizes within species, have different prey preferences and prey species-specific predation behavior [38], as suggested by the species-selective depredation observed in this study. For some odontocetes, predation and other

behaviors may also vary by sympatric populations (e.g., for killer whales in the northeast Pacific Ocean, transients feed on marine mammals and residents target fishes, [56]). Pelagic marine predators' detection, search and attack behavior and preferences for different prey species might be a function of prey species' net energy value (i.e., the prey's caloric value compared to the energy required to catch and handle the prey); and chemical, visual (e.g., size, shape, color and movement), acoustic and textural characteristics [38, 57–62]. However, median length was found in this study to be unrelated to depredation rate. Globally, false killer whales have a broad diet that includes coastal and demersal fishes and epipelagic and oceanicneritic squids, as well as pelagic fishes that are also captured in pelagic longline fisheries, including commercial species such as tunas, dolphinfish, wahoo and sickle pomfret (monchong) [44, 63, 64].

Observers often cannot determine the species of odontocete depredated catch from the small portion of the carcass remaining on the gear. The species-specific odontocete depredation rates for teleosts are conservative underestimates because they do not account for over half of records of odontocete depredated teleosts for which the observer could not identify the catch to the species level. A relatively small proportion of total captured tunas, billfishes and other teleosts lacking odontocete depredated sharks (no rays were odontocete depredated) to the species level while some captured elasmobranchs without odontocete depredation were not identified to the species level, and therefore the elasmobranch species-specific depredation rates may be conservative overestimates. The observer program database also lacks records of pre-catch losses, including when odontocetes and other predators completely remove catch from the gear prior to gear retrieval, as well as catch that falls from the gear due to mechanical action, representing an additional data limitation that affects the accuracy of predicted species-specific odontocete depredation rates.

More robust estimates of species-specific odontocete depredation rates could be derived through research using observer data from longline fisheries that standardize fishing effort to account for potentially informative predictors of odontocete depredation. As conduced previously to explore a response of odontocete depredation of combined species of catch, models with a response of species-specific depredation rate could include predictors such as unique vessel, consecutive sets within a trip, season, geospatial location, hooks per set, mainline length, soak duration, bathymetry and sea surface temperature [26, 32, 37, 45].

While a rare event, odontocete depredation may cause substantial economic and operational costs in the Hawaii tuna longline fishery. This includes direct costs from lost catch [16, 65]. For example, Fader et al. [16] estimated that in the Hawaii tuna longline fishery, odontocete depredation of bigeye and yellowfin tunas and dolphinfish, the three most frequently retained species in this fishery, had an annual value of USD 1 million. Other economic costs result from odontocete depredation of bait, lost and damaged fishing gear and time for crew to repair and replace it, lost fishing time when vessels move or wait to make another set after observing depredation, and increased fishing effort to make up for lost catch [15, 16].

Depredation also leads to bycatch fishing mortality and sublethal effects such as altered foraging behavior, distributions, diet and demographics [7, 17–19]. The false killer whale is the main odontocete species captured in the Hawaii fishery. During the study period (2003–2023) about 70% of captured marine mammals were false killer whales (of 173 odontocetes identified to the species level or higher taxonomic group that could be differentiated from false killer whales). The main Hawaiian Islands insular false killer whale population segment is listed as endangered under the U.S. Endangered Species Act [66], while trends in the abundance of other false killer whale population segments that are exposed to the Hawaii longline and other fisheries are highly uncertain [67]. There are no known odontocete depredation and bycatch mitigation methods for pelagic longline fisheries across the tiers of a sequential mitigation hierarchy that are effective, do not harm odontocetes, and are likely to have acceptable costs on economic viability, practicality and crew safety [15, 21, 24, 25, 45]. To date, measures recommended by an advisory team formed under the U.S. Marine Mammal Protection Act and interventions adopted by the national management authority (weak circle hooks, permanent static closed area, fleetwide bycatch cap that triggers an area closure, and prescribed handling and release practices) have been unsuccessful in meeting management objectives [22].

Study findings might provide evidence to support the identification and innovation of effective and commercially viable methods to mitigate odontocete fisheries depredation and bycatch risk. For example, the spatial and temporal distribution of catch rates of species with and without relatively high odontocete depredation rates could be explored. Fishing effort could be conducted in areas and periods with relatively low catch rates of species with low commercial value and with high odontocete depredation rates that maintains economic viability but lowers odontocete depredation and catch rates [68, 69]. Managing the fishing depth and time of day of fishing might similarly enable reducing catch rates of species with high odontocete depredation rates that also have low commercial value [16, 70]. These findings on odontocete depredation strategies might also contribute to the development of passive acoustic decoys and deterrents. Incorporating objects into the fishing gear may act as decoys if they effectively simulate the acoustic target strength (TS) of the main depredated catch specieshence confusing or frustrating depredating whales [24, 37]. Or, if the objects mimic the TS of a species that odontocetes avoid depredating, then the object may effectively deter depredation behavior [37], such as sharks, which lack swimbladders and have a low TS relative to teleosts. For species with swimbladders, the swimbladder accounts for most (>90%) of fishes' acoustic backscatter [71, 72]. Similarly, hollow alloy spheres attached near terminal tackle have been proposed to reduce odontocete-longline depredation where the object might cause odontocetes to perceive the prev as unusual or the sphere might interfere with their echolocation [73]. This expanded evidence of the species-selective depredation strategies of odontocetes that are exposed to fisheries bycatch promises to contribute to mitigating this human-wildlife conflict.

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#### References

- 1. Bowen WD. Role of marine mammals in aquatic ecosystems. Mar Ecol Prog Ser. 1997; 158: 267–274.
- Roman J, Estes JA, Smith C, Costa D, McCarthy J, Nation JB, et al. Whales as marine ecosystem engineers. Front Ecol Environ. 2014; 12: 377–385.

- Doughty CE, Roman J, Faurby S, Wolf A, Haque A., Bakker E, et al. Global nutrient transport in a world of giants. Proc Natl Acad Sci USA. 2016; 113: 868–873. <u>https://doi.org/10.1073/pnas.1502549112</u> PMID: 26504209
- Estes JA, Doak DF, Springer A., Williams T. Causes and consequences of marine mammal population declines in southwest Alaska: A food-web perspective. Philos Trans R Soc Lond B Biol Sci. 2009; 364: 1647–1658. https://doi.org/10.1098/rstb.2008.0231 PMID: 19451116
- Estes JA, Heithaus M, McCauley DJ, Rasher D, Worm B. Megafaunal impacts on structure and function of ocean ecosystems. Annu Rev Environ Resour. 2016; 41: 83–116.
- Kiszka J, Woodstock M, Heighaus M. Functional roles and ecological importance of small cetaceans in aquatic ecosystems. Front. Mar. Sci. 2022; https://doi.org/10.3389/fmars.2022.803173
- Read AJ. The looming crisis: Interactions between marine mammals and fisheries. J Mammal. 2008; 89: 541–548.
- Davidson AD, Boyer A, Kim H, Pompa-Mansilla S, Hamilton M, Costa D, et al. Drivers and hotspots of extinction risk in marine mammals. Proc Natl Acad Sci USA. 2012; 109: 3395–3400. https://doi.org/10. 1073/pnas.1121469109 PMID: 22308490
- 9. Nelms S, Alfaro-Shigueto J, Arnould J, Avila I, Nash S, Campbell E, et al. Marine mammal conservation: Over the horizon. Endanger Species Res. 2021; 44:291–325.
- Wallace B, Kor C, Dimatteo A, Lee T, Crowder L, Lewison R. Impacts of fisheries bycatch on marine turtle populations worldwide: toward conservation and research priorities. Ecosphere. 2013; 4: 1–49.
- Dias M, Martin R, Pearmain E, Burfield I, Small C, Phillips R, et al. Threats to seabirds: A global assessment. Biol Conserv. 2019; 237: 525–537.
- Dulvy NK, Pacoureau N, Rigby CL, Pollom R, Jabado R Ebert D, et al. Overfishing drives over one third of all sharks and rays toward a global extinction crisis. Curr Biol. 2021; 31: 4773–4787. <u>https://doi.org/ 10.1016/j.cub.2021.08.062</u> PMID: 34492229
- Turvey ST, Pitman RL, Taylor BL, Barlow J, Akamatsu T, Barrett LA, et al. First human-caused extinction of a cetacean species? Biol Lett. 2007; 3: 537–540. <u>https://doi.org/10.1098/rsbl.2007.0292</u> PMID: 17686754
- 14. Avila IC, Kaschner K, Dormann CF. Current global risks to marine mammals: taking stock of the threats. Biol Conserv. 2018; 221: 44–58.
- Gilman E, Brothers N, McPherson G, Dalzell P. Review of cetacean interactions with longline gear. J Cetacean Res Manag. 2006; 8: 215–223.
- Fader J, Marchetti J, Schick R, Read A. No free lunch: Estimating the biomass and ex-vessel value of target catch lost to depredation by odontocetes in the Hawaii longline tuna fishery. Can J Fish Aquat Sci. 2023; https://doi.org/10.1139/cjfas-2022-0156
- 17. Hall MA, Alverson D, Metuzals K. By-catch: problems and solutions. Mar Poll Bull. 2000; 41: 204–19.
- Tixier P, Barbraud C, Pardo D, Gasco N, Duhamel G, Guinet C. Demographic consequences of fisheries interaction within a killer whale (Orcinus orca) population. Mar Biol. 2017; 164:170.
- Tixier P, Lea M, Hindell M, Welsford D, Mazé C, Gourguet S, et al. When large marine predators feed on fisheries catches: global patterns of the depredation conflict and directions for coexistence. Fish Fish. 2021; 22: 31–53.
- **20.** Hamer DJ, Childerhouse S, Gales NJ. Odontocete bycatch and depredation in longline fisheries: a review of available literature and of potential solutions. Mar Mam Sci. 2012; 28: E345–E374.
- 21. Werner T, Northridge S, Press K, Young N. Mitigating bycatch and depredation of marine mammals in longline fisheries. ICES J Mar Sci. 2015; 72: 1576–1586.
- 22. Fader J, Elliott B, Read A. The challenges of managing depredation and bycatch of toothed whales in pelagic longline fisheries: Two U.S. case studies. Front Mar Sci. 2021; https://doi.org/10.3389/fmars. 2021.618031
- NMFS. Taking of Marine Mammals Incidental to Commercial Fishing Operations; False Killer Whale Take Reduction Plan. Fed Regist. 2012; 77: 71259.
- 24. FAO. Report of the Expert Workshop on Means and Methods for Reducing Marine Mammal Mortality in Fishing and Aquaculture Operations. FAO Fisheries and Aquaculture Report No.1231. 2018; Food and Agriculture Organization of the United Nations, Rome.
- Hamilton S, Baker GB. Technical mitigation to reduce marine mammal bycatch and entanglement in commercial fishing gear: lessons learnt and future directions. Rev Fish Biol Fish. 2019; 29: 223–247.
- Forney KA, Kobayashi DR, Johnston D, Marchetti J, Marsik MG. What's the catch? Patterns of cetacean bycatch and depredation in Hawaii-based pelagic longline fisheries. Mar Ecol. 2011; 32: 380–391.

- Hazen EL, Scales KL, Maxwell SM, Briscoe DK, Welch H, Bograd S, et al. A dynamic ocean management tool to reduce by-catch and support sustainable fisheries. Sci Adv 2018; <a href="https://doi.org/10.1126/sciadv.aar3001">https://doi.org/10.1126/sciadv.aar3001</a> PMID: 29854945
- La Manna G., Arrostuto N, Campisi S, Manghi M, Fois N, Ceccherelli G. Acoustic detection of bottlenose dolphin depredation on nets and implications for conservation. Aquat Conserv. 2023; 33: 179–190.
- Gilman E, Chaloupka M, Bellquist L, Bowlby H, Taylor N. Individual and fleetwide bycatch thresholds in regional fisheries management frameworks. Rev Fish Biol Fish. 2023; 34: 253–270.
- Moreno CA, Castro R, Mújica L, Reyes P. Significant conservation benefits obtained from the use of a new fishing gear in the Chilean Patagonian toothfish fishery. CCAMLR Science. 2008; 15: 79–91.
- Rabearisoa N, Bach P, Tixier P, Guinet C. Pelagic longline fishing trials to shape a mitigation device of the depredation by toothed whales. J Exp Mar Biol Ecol. 2012;432–433: 55–63.
- Garrison LP. Interactions between marine mammals and pelagic longline fishing gear in the U.S. Atlantic Ocean between 1992 and 2004. Fish Bull. 2007; 105: 408–417.
- Donoghue M, Reeves R, Stone G. (eds.). Report of the Workshop on Interactions Between Cetaceans and Longline Fisheries. New England Aquarium Aquatic Forum Series Report. 03–1. 2002; New England Aquarium Press, Boston.
- Doksæter L, Godø OR, Handegard NO, Kvadsheim PH, Lam F., Donovan C, et al. Behavioral responses of herring (Clupea harengus) to 1–2 and 6–7 kHz sonar signals and killer whale feeding sounds. J Acoust Soc Am. 2009; 125: 554–564. https://doi.org/10.1121/1.3021301 PMID: 19173441
- Mooney TA, Pacini A, Nachtigall PE. False killer whale (Pseudorca crassidens) echolocation and acoustic disruption: implications for longline bycatch and depredation. Can J Zool. 2009; 87: 726–733.
- **36.** McPherson G, Nishida T. An Overview of Toothed Whale Depredation Mitigation Efforts in the Indo-Pacific Region. 2010; Secretariat of the Pacific Community, Noumea, New Caledonia.
- O'Connell V, Straley J, Liddle J, Wild L, Behnken L, Falvey D, et al. Testing a passive deterrent on longline to reduce sperm whale depredation in the Gulf of Alaska. ICES J Mar Sci. 2015; 72: 1667–1672.
- 38. Gilman E, Chaloupka M, Bach P, Fennell H, Hall M, Musyl M, et al. Effect of pelagic longline bait type on species selectivity: A global synthesis of evidence. Rev Fish Biol Fish. 2020; 30: 535–551.
- Bayse SM, Kerstetter D. Assessing bycatch reduction potential of variable strength hooks for pilot whales in a western north Atlantic pelagic longline fishery. J N C Acad Sci. 2010; 126: 6–14.
- **40.** Bigelow K, Kerstetter D, Dancho M, Marchetti J. Catch rates with variable strength circle hooks in the Hawaii-based tuna longline fishery. Bull Mar Sci. 2012; 88: 425–447.
- McLellan W, Arthur L, Mallette S, Thornton S, McAlarney R, Read A, et al. Longline hook testing in the mouths of pelagic odontocetes. ICES J Mar Sci. 2015; 72: 1706–1713.
- Gearin PJ, Pfeifer R, Jeffries SJ, DeLong RL, Johnson M. Results of the 1986–87 California sea lionsteelhead trout predation control program at the Hiram M. Chittenden Locks. NWAFC Processed Report 88–30. 1988; Alaska Fisheries Science Center, National Marine Fisheries Service, Seattle.
- Andersen M, Forney K, Cole T, Eagle T, Angliss R, Long K, et al. Differentiating Serious and Non-Serious Injury of Marine Mammals: Report of the Serious Injury Technical Workshop, 10–13, September 2007. NOAA Technical Memorandum NMFS-OPR-39. 2008; National Marine Fisheries Service, Seattle.
- 44. Oleson E, Boggs C, Forney K, Hanson M, Kobayashi D, Taylor B, et al. Status Review of Hawaiian Insular False Killer Whales (Pseudorca crassidens) under the Endangered Species Act. 2010; Pacific Islands Fisheries Science Center, National Marine Fisheries Service, Honolulu.
- 45. Fader JE, Baird R, Bradford A, Dunn D, Forney K, Read A. Patterns of depredation in the Hawaii deepset longline fishery informed by fishery and false killer whale behavior. Ecosphere 2021; do: https://doi. org/10.1002/ecs2.3682
- NMFS. Hawaii Longline Observer Program Field Manual. Version LM.17.02. 2017; Pacific Islands Regional Office, National Marine Fisheries Service, Honolulu.
- Tuyl F, Gerlach R, Mengersen K. Comparison of Bayes-Laplace, Jeffreys, and other priors: The case of zero events. Am Stat. 2008; 62: 40–44.
- Lin L, Chu H. Meta-analysis of proportions using generalized linear mixed models. Epidemiology. 2020; 31: 713–717. https://doi.org/10.1097/EDE.000000000001232 PMID: 32657954
- 49. Gilman E, Chaloupka M, Benaka L, Bowlby H, Fitchett M, Michel Kaiser M, et al. Phylogeny explains capture mortality of sharks and rays in pelagic longline fisheries: a global meta-analytic synthesis. Sci Rep. 2022; https://doi.org/10.1038/s41598-022-21976-w PMID: 36307432
- Dorai-Raj S. binom: Binomial confidence intervals for several parameterizations. R package version 1.1–1.1. 2022; https://CRAN.R-project.org/package=binom.

- Roda M, Gilman E, Huntington T, Kennelly S, Suuronen P, Chaloupka M, et al. A Third Assessment of Global Marine Fisheries Discards. FAO Fisheries and Aquaculture Technical Paper 633. ISBN 978-92-5-131226-1. 2019; Food and Agriculture Organization of the United Nations, Rome.
- 52. Puth M, Neuhäuser M, Ruxton G. Effective use of Pearson's product–moment correlation coefficient. Anim Behav. 2014; 93: 183–189.
- Wei T, Viliam Simko V. R package 'corrplot': Visualization of a correlation Mmtrix (Version 0.92). 2021; https://github.com/taiyun/corrplot.
- 54. Agresti A. Categorical data analysis (second edition). 2002; Wiley: New York.
- Aragon T. epitools: Epidemiology Tools. R package version 0.5–10.1. 2020; <u>https://cran.r-project.org/web/packages/epitools/index.html</u>.
- 56. Saulitis E, Matkin C, Barrett-Lennard L, Heise K, Ellis G. Foraging strategies of sympatric killer whale (Ocrinus orca) populations in Prince William Sound, Alaska. Mar Mamm Sci. 2000; 16: 94–109.
- 57. Hart P. Foraging in teleost fishes. pp. 211–235 IN Pitcher T (Ed). Behaviour of Teleost Fishes, 2nd ed. 1993; Chapman & Hall, London.
- Krivan V. Optimal foraging and predator-prey dynamics. Theor Popul Biol. 1996; 49: 265–290. <a href="https://doi.org/10.1006/tpbi.1996.0014">https://doi.org/10.1006/tpbi.1996.0014</a> PMID: 8813025
- Ward P. Empirical estimates of historical variations in the catchability and fishing power of pelagic longline fishing gear. Rev Fish Biol Fish. 2008; 18: 409–426.
- Yokota K, Kiyota M, Okamura H. Effect of bait species and color on sea turtle bycatch in a pelagic longline fishery. Fish Res. 2009; 97: 53–58.
- Lokkeborg S, Siikavuopio S, Humborstad O, Utne-Palm A, Ferter K. Toward more efficient longline fisheries: Fish feeding behavious, bait characteristics and development of alternative baits. Rev Fish Biol Fish. 2014; 24:985–1003.
- Piovano S, Farcomeni A, Giacoma C. Effects of chemicals from longline baits on the biting behaviour of loggerhead sea turtles. Afr J Mar Sci. 2012; 34: 1–5.
- Baird RW, Gorgone A, McSweeney DJ, Webster D, Salden D, Deakos M, et al. False killer whales (Pseudorca crassidens) around the main Hawaiian Islands: Long-term site fidelity, inter-island movements, and association patterns. Mar Mammal Sci. 2008; 24: 591–612.
- Riccialdelli L, Goodall N. Intra-specific trophic variation in false killer whales (Pseudorca crassidens) from the southwestern South Atlantic Ocean through stable isotopes analysis. Mamm Biol. 2015; 80: 298–302.
- Peterson MJ, Mueter F, Criddle K, Haynie A. Killer whale depredation and associated costs to Alaskan sablefish, Pacific halibut and Greenland turbot longliners. PLoS ONE. 2014; <u>https://doi.org/10.1371/journal.pone.0088906 PMID: 24558446</u>
- 66. NMFS. Main Hawaiian Islands Insular False Killer Whale (*Pseudorca crassidens*) Distinct Population Segment 5-Year Review: Summary and Evaluation. 2022; Pacific Islands Regional Office, National Marine Fisheries Service, Honolulu.
- 67. Carretta J, Oleson E, Forney K, Weller D, Lang A, Baker J, et al. U.S. Pacific Marine Mammal Stock Assessments: 2022. NOAA Technical Memorandum NOAA-TM-NMFS-SWFSC-684. 2023; Southwest Fisheries Science Center, National Marine Fisheries Service, La Jolla, California, USA.
- Slooten E. Effectiveness of area-based management in reducing bycatch of the New Zealand Dolphin. Endanger Species Res. 2013; 20: 121–130.
- 69. FAO. Report of the Expert Meeting on Other Effective Area-Based Conservation Measures in the Marine Capture Fishery Sector. FAO Fisheries and Aquaculture Report No. 1301. 2019; Food and Agriculture Organization of the United Nations, Rome.
- Musyl M, Brill R, Curran D, Fragoso N, McNaughton L, Nielsen A, et al. Postrelease survival, vertical and horizontal movements, and thermal habitats of five species of pelagic sharks in the central Pacific Ocean. Fish Bull. 2011; 109: 341–361.
- **71.** Foote KG. Importance of the swimbladder in acoustic scattering by fish: A comparison of gadoid and mackerel target strengths. J Acoust Soc Am. 1980; 67: 2084–2089.
- 72. Bertrand A, Josse E. Tuna target-strength related to fish length and swimbladder volume. ICES J Mar Sci. 2000; 57: 1143–1146.
- Deveau D, McPherson G. Application of passive acoustic reflectors to mitigate toothed whale depredation on longlines. J Acoust Soc Am. 2011; https://doi.org/10.1121/1.3587806