

IDENTIFYING DATA GAPS AND OPPORTUNITIES FOR UPDATING M ORPHOM ETRIC RELATIONSHIPS AND COLLECTING BIOLOGICAL SAM PLES FOR PRIORITY SPECIES IN EASTERN PACIFIC OCEAN TUNA FISHERIES (SAC-14 INF-J)

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## Outline

- Background
- Objectives
- EPO data gaps - M orphometric relationships

B e.g., length-weight (L-W), length-length (L-L), weight-weight (W-W)

- EPO data gaps - Biological sampling

B e.g., tissues, stomachs, vertebral centra, otoliths, gonads

- Potential sampling opportunities
- Final considerations



## Background: M orphometric relationships

- M orphometric data are critical to several research and reporting activities
ß Stock assessments
B Ecological assessments (e.g., EASI-Fish)
- Relationships vary by species, region, year
- Species and size composition of catches differs by fishing gear
- Variability may influence assessments and increase uncertainty
- Catch estimations are influenced by morphometric relationships
ß L-W data are used to convert catch data in numbers to weights and vice versa
B W-W data required to convert processed weight to whole weight



## Background: M orphometric relationships

- Various weight metrics
ß whole weight
B gilled and gutted weight
B headed, tailed and gutted weight
- Various length metrics

B fork length
B total length
B lower-jaw fork length (billfish)


B eye-fork length (billfish)
B precaudal length and interdorsal length (sharks)

- Improvements to morphometric relationships are essential for improving precision


## Background: Biological sampling

- Biological data needed to parameterize stock assessment models e.g.,

B stock structure (e.g., tagging and genetics)
B growth and longevity of tunas (e.g., otoliths and tagging)
B reproductive biology (e.g., histology)
B natural mortality (M) (e.g., tagging and growth parameters for $M$ estimators)
B movement (e.g., conventional and electronic tagging)


## Background: Biological sampling

- Biological data needed to parameterize ecological assessments e.g.,

B to characterize age, growth and reproduction (i.e., productivity component of EASI-Fish)
B stomach content data (foundation of ecosystem model)
B experiments on consumption rates ( $\mathrm{Q} / \mathrm{B}$ ratio parameter in ecosystem models)
B genetic information (e.g., CKM R for sharks, SAC-12-14)


## Initiation of Project F.3.a.

- Staff initiated Project F.3.a. to address data gaps and evaluate sampling feasibility

B morphometric relationships
B biological sampling

- Similar work conducted in WCPO (Project 90: SC18-ST-IP-04)

B SPC Oceanic Fisheries Programme established PS and LL observer measurement protocols
B developed measurement guidelines and forms (GEN-4 Conversion Factor)
B record multiple measurements on the same fish (e.g., FL, TL, whole weight, processed weight)
B purpose: to build comprehensive database of various length and weight types
B database allows scientists to develop the morphometric relationship needed for assessments
B established Pacific M arine Specimen Bank (biological samples)

- SAC-14 INF-J summarizes staff discussions on Project F.3.a

B complementary to documents on data improvements (SAC-12-09, WSDAT-01-01, WSDAT-01-Report, SAC14 INF-M, SAC-14 INF-L)
B creating awareness of data deficiencies
B seeking ways to improve data collection

## Objectives outlined in SAC-14 INF-J

## - Objectives

B identify data gaps in morphometric relationships and biological sampling
B identify potential opportunities for morphometrics and opportunistic biological sampling (for tunas, billfishes, prioritized species)
B provide considerations that will determine the success of the project (F.3.a)
B improve data collection to better align with scientific research under the Antigua Convention


## Data gaps: M orphometric relationships $\mathrm{W}=\mathrm{a} \mathrm{L}^{\mathrm{b}}$

- Relationships for tropical tunas are outdated by several decades
ß YFT (Wild 1986)
ß BET (Nakamura and Uchiyama 1966)
B SKJ (Hennemuth 1959)
B not representative of current EPO populations and fisheries
$ß$ there is indication of spatial variation in average sizes, temporal variation is expected
B critical for improving estimation of annual catches for all fisheries
B measurements of lengths and weights pre- and post-processing are required



## Data gaps: M orphometric relationships

- Staff derive estimates of annual and total catch of tunas, by species, using

B observer records, vessel logbooks, cannery records, port-sampling data (e.g., see WSBET-02-06)

- Data reported are not standardized

B processed or whole weight and/or numbers and lengths
B processing variability between fleets
B some fishers remove operculum and tail, freeze fish in ultra-low temperatures (Langley et al. 2006)
B others chill fish and land fresh fish with only viscera and gills removed (Langley et al. 2006)

- No EPO-specific conversion factors for gilled and gutted weight (SAC-07-04a)

B stock assessment team use conversion factors for entire Pacific Ocean (Langley et al. 2006)

- Current catch estimates may be biased, due to:

B outdated morphometric relationships used to convert from length to weight
B processing methods (e.g., sampling frozen tunas vs. fresh tunas)
$B$ individual variability in the relationships not accounted for (important when fitting models to weight frequencies)

## Data gaps: M orphometric relationships

- Relationships for non-target species are: (see IATTC Special Report 25, SAC-13-11, SAC-09-12)

B non-existent
B outdated
B borrowed from similar species within the region
B based on data from other ocean basins
B do not represent EPO populations

## Data gaps: M orphometric relationships

- EASI-Fish (e.g., SAC-14-12) \& estimates of artisanal shark catches (SAC-14 INF-L):

B different forms of L-W \& W-W relationships available (literature review)
B length measurements (e.g., type: PCL, FL, TL)
B weight measurements (e.g., processed weight, whole weight)
B $W=a L^{b}$ (e.g., EASI-Fish uses TL in cm; $W$ in kg )
ß conversions needed to raise processed weights to whole weights (SAC-14 INF-L)
B analyst must convert to the appropriate form


Photos from Project C.4.b. Long-term sampling Program for catches of sharks in artisanal fisheries in Central America

## Data gaps: Biological sampling

- Routine biological sampling is not conducted
- Routine sampling provides a means for monitoring fishing impacts

B decrease in YFT size at maturity (Schaefer and Fuller 2022; Schaefer 1998)
B such changes can impact productivity, subsequent stock status and management advice



## Data gaps: Biological sampling

- Sampling has been limited to dedicated projects

B M iss changes in biological processes across dynamic conditions (e.g., ENSO events)

- Sampling has not kept pace with modern techniques

B e.g., tissue collection for genetic studies on stock structure and CKM R (sharks) SAC-12-14

- ETP ecosystem model based on antiquated stomach contents (1990s)

B ecological impacts include changes in feeding dynamics



## Identification of sampling opportunities: Overview

- Size composition of catches varies by gear type

B aim to sample across gear and fleet types
B aim to maximize spatial range (coastal-offshore) \& size distributions of fish (juveniles-adults)

- Propose hierarchical sampling approach

B feasibility study (Phase 1)
B pilot study (Phase 2)
B EPO-wide, statistically-robust sampling (Phase 3)

- Aim to collect different length and weight metrics on the same individuals

B allows for comprehensive database for development of conversions \& to account for individual variability

- Opportunistically collect biological samples




## Identification of sampling opportunities: Feasibility study

| Phase 1 | Action | Outcome | Preliminary timeline | Collaborators |
| :---: | :---: | :---: | :---: | :---: |
| $\begin{aligned} & \text { Feasibility } \\ & \text { (Part 1, planning) } \end{aligned}$ | Identify measurements to be taken and biological samples to be collected | List of morphological measurements (e.g., FL, TL, WW, GGW); List of biological samples (e.g., tissues, stomachs) | January-M ay 2024 | Stock assessment, Biology and life-history, Ecosystem and bycatch, and Data Programs; CPCs, fishing industry, SPC-WCPFC |
|  | Identify priority species through literature review and meta-analysis | List of priority species to sample (e.g., silky sharks, hammerhead sharks) |  |  |
|  | Design feasibility studies with sampling protocols for both PS and LL fisheries | Development of data collection forms and data/sample storage protocols |  |  |
|  | $\begin{aligned} & \text { Identify capacity building } \\ & \text { opportunities and potential } \\ & \text { collaborators } \\ & \hline \end{aligned}$ | List of vessels to be used for sampling; List of external collaborators |  |  |
|  | Identify storage opportunities for biological samples | List of potential storage facilities |  |  |
|  | Preliminary design of a database for morphometric measurements and biological samples | Beta database structure developed |  |  |

## Identification of sampling opportunities: Feasibility study

| Phase 1 | Action | Outcome | Preliminary timeline | Collaborators |
| :---: | :---: | :---: | :---: | :---: |
| Feasibility (Part 2, implementation) | IATTC staff to execute feasibility studies aboard class 6 PS and coastal LL tuna fishing vessels | Evaluation of collected data and samples; Revision of sampling protocols prior to implementing pilot phase | June 2024-M ay 2025 | Stock assessment, Biology and life-history, Ecosystem and bycatch, and Data Programs; CPCs, fishing |
|  | Pursue capacity building opportunities with potential collaborators within distant water LL fleets (in preparation for Phase 2, Pilot study) | List of potential distantwater LL fishing vessels for sampling |  | industry, SPC-WCPFC |
|  | Collaborate with statisticians to develop statistically robust sampling design for industrial fisheries (in preparation for Phase 2, Pilot study) | Development of sampling protocol for upscaling sampling to additional vessels in Phase 2, Pilot study |  |  |

## Identification of sampling opportunities: Pilot study

| Phase 2 | Action | Outcome | Preliminary timeline | Collaborators |
| :---: | :---: | :---: | :---: | :---: |
| Pilot sampling | Through collaborations, implement pilot study following lesson's learned and sampling design from Phase 1. Sample across all PS (i.e., class 1-6) vessels, coastal States LL tuna vessels, and distant-water LL vessels. Revise the sampling design as needed. Coordinate logistics for storing samples. | Development of sampling protocols for industrial fisheries; documentation of lesson's learned from Phase 1 and Phase 2 industrial fisheries. Compilation of a dataset to derive L-W relationships for tunas and prioritized species from industrial tuna fisheries; Collection and storage of biological samples (tissues, stomachs, gonads, otoliths, vertebrae) for tunas and priority species | June 2025-M ay 2026 | Stock assessment, Biology and lifehistory, Ecosystem and bycatch, and Data Programs; CPCs, fishing industry, SPCWCPFC |
|  | Pursue collaborations within coastal, multi-gear/multi-species fisheries; Work with statisticians to develop a sampling design for small coastal, multigear fisheries | Development of sampling protocols for coastal-multi-gear fisheries; documentation of lesson's learned from sampling these fisheries |  |  |
|  | Implement sampling in small coastal fisheries. Revise the sampling design as needed. Coordinate logistics for storing samples. | Compilation of a data set to derive L-W relationships for tunas and prioritized species from coastal multi-gear fisheries; Collection and storage of biological samples (tissues, stomachs, gonads, otoliths, vertebrae) for tunas and priority species |  |  |

## Identification of sampling opportunities: EPO-wide study

| Phase 3 | Action | Outcome | Preliminary timeline | Collaborators |
| :---: | :---: | :---: | :---: | :---: |
| EPO-wide, statistically robust sampling | Expand sampling to additional vessels and areas across the EPO as is feasible. Continue sampling on PS class 1-6 vessels, coastal States LL tuna fisheries, distant-water LL fisheries, and coastal multi-gear fisheries | Collection of a robust data set to derive L-W relationships throughout the operational range of EPO fisheries. Store biological samples (tissues, stomachs, gonads, otoliths, vertebrae) for tunas and priority species. | J anuary 2026-M ay 2030 | Stock assessment, Biology and life-history, Ecosystem and bycatch, and Data Programs; CPCs, fishing industry, SPC-WCPFC |
| Establishment of an EPO-wide morphometric and biological database for various fisheries | Analyze data; develop morphometric relationships and conversion factors. Process prioritized biological samples in-house and through collaborations | Development of EPO-wide database of morphometrics and biological data. Biological material storage. Publications of meta-data and morphometric relationships. Projectspecific analysis of biological samples (e.g., stock assessments and ecological assessments) | January 2026-December 2030 | Stock assessment, Ecosystem and bycatch, Biology and life-history, and Data Programs |

## Final Considerations

- Success of the proposed sampling program predicated on collaborations, logistics and funding
- Therefore, staff has designed an iterative approach to the proposed project

B phased approach to upscaling sampling to additional vessels and areas
A biological sampling may occur opportunistically
B noting NOAA's Climate Prediction Center predicts transition to El Niño

- Project is complementary to others on data improvement

B e.g., SAC-12-09, WSDAT-01-01, WSDAT-01-Report, SAC-14 INF-M, SAC-14 INF-L



Questions

## PS sampling protocols: SPC-OFP 2021, GEN-4 Conversion Factor

## Weights and measurements collected

| DETAILS OF WEIGHTS AND MEASUREMENTS COLLECTED |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
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| SETNO. | SHIP'S | LABEL | SPECIES | LENGTHS (in cm.) |  |  |  |  |  | WEIGHTS (in kg.) |  |  |  |  | PROCESSED WGT. |  | LANDED WEIGHT |  | COMMENTS |
|  | TIME | NO. | CODE | UF | US | LF | PF | PS | TL | WHOLE | HEAD | TAIL | GUTS | WET FIN | (kg.) | CODE | (kg.) | CODE |  |
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